Restriction Endonuclease

SibEnzyme®

BamH I

Recognition Sequence:

G↓GATCC
CCTAG↑G

XS

E949m

2000 units

20,000 u/ml

Lot:

Exp:

Store at -20C

SE-Buffers	B	G	O	W	Y	ROSE
%Activity	25-50	25-50	10-25	10-25	100	40

37°C | NO | Y | λ | RR | BSA

For more details scen the code

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CERTIFICATE OF ANALYSIS

Source: *An E.coli strain that carries the cloned BamH I gene from Bacillus amyloliquefaciens H.*

Supplied in:
10 mM Tris-HCl (pH 7.4), 50 mM KCl, 0.1 mM EDTA, 1 mM DTT, 200 µg/ml BSA, 50% glycerol.

Reaction Conditions:
1X SE-Buffer Y, BSA (100 µg/ml). Incubate at 37° C.

1X SE-Buffer Y (pH 7.9 @ 25° C):
33 mM Tris-Ac 66mM KCl
10 mM MgAc 1 mM DTT

Heat Inactivation:
NO (80 °C for 20 minutes).

Unit Definition: One unit is defined as the amount of enzyme required to digest 1 µg of Lambda DNA in 1 hour at 37° C in a total reaction volume of 50 µl.
To obtain 100% activity, BSA should be added to the 1 x reaction mix to a final concentration of 100 µg/ml.

Quality Control Assays
Ligation: After 20-fold overdigestion with BamH I, ~90% of the DNA fragments can be ligated with high -activity T4 DNA Ligase and recut.

16-Hour Incubation: A 50 µl reaction containing in 1 µg of DNA and 40 Units of enzyme incubated for 16 hours resulted in the same pattern of DNA bands as a reaction incubated for 1 hour.

Oligonucleotide Assay: No detectable degradation of a single-stranded and double-stranded oligonucleotide was observed after incubation with 20 units of restriction endonuclease for 3 hours.

Enzyme Properties:
When using a buffer other than the optimal (Supplied) SE-Buffer, it may be necessary to add more enzymes to achieve complete digestion.

Reagents Supplied with Enzyme:
10X SE Buffer Y, BSA (10 mg/ml).