



EcoR I

Recognition Sequence:

E939m XS

5,000 units 20.000 u/ml

GLAATTC. CTTAA†G

Lot: Exp:

Store at -20C

SE-Buffers	В	G	0	w	Υ	ROSE
%Activity	10-25	100	0-10	0-10	100	40
37°C 65°C Υ λ RR BSA						

For more details scen the code



CERTIFICATE OF ANALYSIS

Source: An E.coli strain that carries the cloned EcoR I aene from Escherichia coli.

Supplied in:

10 mM KPO4, 300 mM NaCl, 0.1 mM EDTA, 1 mM DTT, 200 μg/ml BSA, 50% glycerol. 0.15% Triton X-100, pH7.0 @ 25 °C.

Reaction Conditions:

 $1 \times SE$ -Buffer Y, BSA (100 μ g/ml). Incubate at 37 °C.

1X SE-Buffer Y (pH 7.9 @ 25° C): 33 mM Tris-HCl 66mM KAc 10 mM MgAc 1 mM DTT

Heat Inactivation:

Enzyme is inactivated by incubation at 65°C for 20 minutes.

Unit Definition: One unit is defined as the amount of enzyme required to digest 1 µg of Lambda DNA in 1 hour at 37°C in a total reaction volume of 50 µl. To obtain 100% activity, BSA should be added to the 1 x reaction mix to a final concentration of 100 μg/ml. Do not use BSA for long incubation.

Quality Control Assays

Ligation: After 40-fold overdigestion with EcoR I, ~95% of the DNA fragments can be ligated and recut.

16-Hour Incubation: A 50 μl reaction containing in 1 μg of DNA and 40 Units of enzyme incubated for 16 hours resulted in the same pattern of DNA bands as a reaction incubated for 1 hour.

Oligonucleotide Assay: No detectable degradation of a single-stranded and double-stranded oligonucleotide was observed after incubation with 20 units of restriction endonuclease for 3 hours.

Enzyme Properties:

When using a buffer other than the optimal (Supplied) SE-Buffer, it may be necessary to add more enzymes to achieve complete digestion.

Reagents Supplied with Enzyme: 10X SE Buffer Y, BSA (10 mg/ml).