



EcoR I

Recognition Sequence:

xs E057m

500 units 20,000 u/ml G\$\text{AATTC} CTTAATG

Lot: Exp:

Store at -20C

W V ROSE

 SE-Buffers
 B
 G
 O
 W
 Y
 ROSE

 %Activity
 50-75
 75-100
 75-100
 75-100
 50-75
 50

 37°C
 65°C
 EcoRI
 λ
 RR
 BSA

For more details scen the code

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CERTIFICATE OF ANALYSIS

Source: An E.coli strain that carries the cloned EcoR I gene from Escherichia coli.

Supplied in:

10 mM Tris-HCl (pH 7.5), 200 mM NaCl, 0.1 mM EDTA, 7 mM 2-mercaptoethanol, 100 μg/ml BSA, 50% glycerol.

Reaction Conditions:

 $1\times$ SE-Buffer EcoR I, BSA (100 $\mu g/ml).$ Incubate at 37° C.

1X SE-Buffer EcoR I (pH 7.6 @ 25° C):

 $\begin{array}{ccc} 10~\text{mM Tris-HCl} & 50~\text{mM NaCl} \\ 10~\text{mM MgCl}_2 & 1~\text{mM DTT} \end{array}$

Heat Inactivation:

Enzyme is inactivated by incubation at 65°C for 20 minutes.

<u>Unit Definition</u>:0ne unit is defined as the amount of enzyme required to digest 1 μ g of Lambda DNA in 1 hour at 37° C in a total reaction volume of 50 μ l. To obtain 100% activity, BSA should be added to the 1 x reaction mix to a final concentration of 100 μ g/ml. High enzyme concentration and using of nonoptimal buffer may result in star activity. Do not use BSA for long incubation.

Quality Control Assays

 $\label{light} \frac{Ligation:}{Ligation:} After 40-fold overdigestion with EcoR I, \sim 95\% \\ of the DNA fragments can be ligated and recut.$

<u>16-Hour Incubation</u>: A 50 µl reaction containing in 1 µg of DNA and 40 Units of enzyme incubated for 16 hours resulted in the same pattern of DNA bands as a reaction incubated for 1 hour.

Oligonucleotide Assay: No detectable degradation of a single-stranded and double-stranded oligonucleotide was observed after incubation with 20 units of restriction endonuclease for 3 hours.

Enzyme Properties:

When using a buffer other than the optimal (Supplied) SE-Buffer, it may be necessary to add more enzymes to achieve complete digestion.

Reagents Supplied with Enzyme:

10X SE Buffer EcoR I, BSA (10 mg/ml).