

Restriction
Endonuclease



FauND I

Recognition
Sequence:

CA↓TATG
CTAT↑AC

XS

E009m
500 units
10,000 u/ml

Lot:
Exp:
Store at -20C

SE-Buffers	B	G	O	W	Y	ROSE
%Activity	50-75	75-100	10-25	50-75	100	100

37°C 65°C Y λ RR minimal

For more details
scan the code



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CERTIFICATE OF ANALYSIS

Source: An *E.coli* strain that carries the cloned FauND I gene from *Flavobacterium aquatili* ND.

Supplied in:
10 mM Tris-HCl (pH 7.6), 50 mM KCl, 0.1 mM EDTA,
200 µg/ml BSA, 1 mM DTT, 50% glycerol.

Reaction Conditions:
1x SE-Buffer Y, BSA(100 µg/µl) Incubate at 37° C.

1X SE-Buffer Y (pH 7.9 @ 25° C):
33 mM Tris-Ac 66 mM KAc
10 mM MgAC 1 mM DTT

Heat Inactivation:
Enzyme is inactivated by incubation at 65 °C for 20 minutes.

Unit Definition: One unit is defined as the amount required to digest 1 µg of λ DNA in 1 hour at 37° C in a total reaction volume of 50 µl. To obtain 100% activity, BSA should be added to the 1X reaction mix to final concentration of 100 µg/µl.

Quality Control Assays

Ligation : After 10-fold overdigestion with FauND I, approximately 80% of the DNA fragments can be ligated with T4 DNA Ligase and recut. In the presence of 10% PEG ligation is better.

16-Hour Incubation: A 50 µl reaction containing 1 µg of DNA and 10 Units of enzyme incubated for 16 hours resulted in the same pattern of DNA bands as a reaction incubated for 1 hour.

Oligonucleotide Assay: No detectable degradation of a single-stranded and double-stranded oligonucleotide was observed after incubation with 10 units of restriction endonuclease for 3 hours.

Note:

Sensitive to impurities in some DNA preparations. For example, DNA purified by standard miniprep procedures is cleaved at lower rates.

Enzyme Properties:

When using a buffer other than the optimal (Supplied) SE-Buffer, it may be necessary to add more enzymes to achieve complete digestion.

Reagents Supplied with Enzyme:

10X SE Buffer Y, 100X BSA (10 mg/µl).