

Bpul0 I

Recognition Sequence:

5 E149 200 units 5,000 u/ml CC↓TNAGC GGANT↑CG

Exp:

Store at -20C

 SE-Buffers
 B
 G
 O
 W
 Y
 ROSE

 %Activity
 10-25
 25-50
 50-75
 50-75
 25-50
 100

37°C 80°C Κ λ RR

For more details scen the code

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CERTIFICATE OF ANALYSIS

<u>Source</u>: An E.coli strain that carries recombinant plasmids.

Supplied in:

10 mM Tris-HCl (pH 7.5), 50 mM NaCl, 0.1 mM EDTA, 1 mM DTT, 50% glycerol.

Reaction Conditions:

1X SE-Buffer K. Incubate at 37° C.

Heat Inactivation:

Enzyme is inactivated by incubation at 80°C for 20 minutes.

<u>Unit Definition</u>: One unit is defined as the amount of enzyme required to digest $1 \mu g$ of Lambda DNA in $1 \mu g$ hour at 37° C in a total reaction volume of $50 \mu l$.

Quality Control Assays

<u>Ligation</u>:After 10-fold overdigestion with Bpu10 I, 80% of the DNA fragments can be ligated. Of these 90% can be recut. In the presence of 10% PEG ligation is better.

16-Hour Incubation: A 50 μl reaction containing 1 μg of DNA and 5 Units of enzyme incubated for 16 hours resulted In the same pattern of DNA bands as a reaction

incubated for 1 hour.

High enzyme concentration may result in star activity or incomplete DNA cleavage. We recommend increasing

Oligonucleotide Assay: No detectable degradation of a single-stranded and double-stranded oligonucleotide was observed after incubation with 5 units of restriction endonuclease for 3 hours.

incubation time instead of using an excess of Bpu101.

Enzyme Properties:

When using a buffer other than the optimal (Supplied) SE-Buffer, it may be necessary to add more enzymes to achieve complete digestion.

Reagents Supplied with Enzyme: 10X SF Buffer K.

The minimum number of units that resulted in complete digestion of 1 μg of substrate DNA in 16 hours is 0.5.