

# BamH I

GLGATCC Recognition Sequence: CCTAGTG E022 Lot:

> 20,000 units 20.000 u/ml

Exp: Store at -20C

SE-Buffers ROSE 25-50 100 75-100 75-100 25-50 100

**BSA** 

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# CERTIFICATE OF ANALYSIS

Source: An E.coli strain that carries the cloned BamH I gene from Bacillus amyloliquefaciens H.

## Supplied in:

10 mM Tris-HCl (pH 7.6), 50 mM NaCl, 0.1 mM EDTA, 1 mM DTT, 100 µg/ml BSA, 50% glycerol.

## Reaction Conditions:

1X SE-Buffer G, BSA (100 µg/ml). Incubate at 37° C.

1X SE-Buffer G (pH 7.6 @ 25° C): 10 mM Tris-HCL 50mM NaCl 10 mM MgCl<sub>2</sub> 1 mM DTT

**Heat Inactivation:** 

Enzyme is inactivated by incubation at 65°C for 20 minutes.

Unit Definition: One unit is defined as the amount of enzyme required to digest 1 µg of Lambda DNA in 1

hour at 37°C in a total reaction volume of 50 µl. To obtain 100% activity, BSA should be added to the 1 x reaction mix to a final concentration of 100 μg/ml.

of DNA and 20 Units of enzyme incubated for 16 hours

was observed after incubation with 20 units of restriction

### Quality Control Assays Ligation: After 20-fold overdigestion with BamH I, ~90%

of the DNA fragments can be ligated with high -activity T4 DNA Ligase and recut. 16-Hour Incubation: A 50 µl reaction containing in 1 µg

resulted in the same pattern of DNA bands as a reaction incubated for 1 hour. Do not use BSA for long incubation.

endonuclease for 3 hours.

**Enzyme Properties:** 

When using a buffer other than the optimal (Supplied) SE-Buffer, it may be necessary to add more enzymes to achieve complete digestion.

Reagents Supplied with Enzyme: 10X SE Buffer G. BSA (10 mg/ml).

Oligonucleotide Assay: No detectable degradation of a single-stranded and double-stranded oligonucleotide