

Restriction
Endonuclease



BamH I

Recognition
Sequence:

G↓GATCC
CCTAG↑G

S

E021

4,000 units
20,000 u/ml

Lot:

Exp:

Store at -20C

SE-Buffers	B	G	O	W	Y	ROSE
%Activity	25-50	100	75-100	75-100	25-50	100

37°C

65°C

G

λ

RR

BSA

For more details
scan the code



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CERTIFICATE OF ANALYSIS

Source: An *E.coli* strain that carries the cloned *BamH I* gene from *Bacillus amyloliquefaciens* H.

Supplied in:

10 mM Tris-HCl (pH 7.6), 50 mM NaCl, 0.1 mM EDTA,
1 mM DTT, 100 µg/ml BSA, 50% glycerol.

Reaction Conditions:

1X SE-Buffer G, BSA (100 µg/ml). Incubate at 37° C.

1X SE-Buffer G (pH 7.6 @ 25° C):

10 mM Tris-HCl 50mM NaCl
10 mM MgCl₂ 1 mM DTT

Heat Inactivation:

Enzyme is inactivated by incubation at 65° C for 20 minutes.

Unit Definition: One unit is defined as the amount of enzyme required to digest 1 µg of Lambda DNA in 1 hour at 37° C in a total reaction volume of 50 µl. To obtain 100% activity, BSA should be added to the 1 x reaction mix to a final concentration of 100 µg/ml.

Quality Control Assays

Ligation: After 20-fold overdigestion with BamH I, ~90% of the DNA fragments can be ligated with high -activity T4 DNA Ligase and recut.

16-Hour Incubation: A 50 µl reaction containing in 1 µg of DNA and 20 Units of enzyme incubated for 16 hours resulted in the same pattern of DNA bands as a reaction incubated for 1 hour.

Do not use BSA for long incubation.

Oligonucleotide Assay: No detectable degradation of a single-stranded and double-stranded oligonucleotide was observed after incubation with 20 units of restriction endonuclease for 3 hours.

Enzyme Properties:

When using a buffer other than the optimal (Supplied) SE-Buffer, it may be necessary to add more enzymes to achieve complete digestion.

Reagents Supplied with Enzyme:

10X SE Buffer G, BSA (10 mg/ml).