



Ksp22 I

Recognition Sequence:

1,000 units 20.000 u/ml

E081

TIGATCA **ACTAG** † T

Lot: Exp:

Store at -20°C

SE-Buffers W ROSE 50-75 75-100 50-75 50-75 25-50 100 **BSA**



Ph/F+7(383)333-6853 info@sibenzyme.com www.sibenzvme.com

CERTIFICATE OF ANALYSIS

Source: Kurthia species 22.

Supplied in:

10 mM Tris-HCl (pH 7.5), 250 mM NaCl, 0.1 mM EDTA, 7 mM 2-mercaptoethanol, 200 µg/ml BSA, 50% glycerol.

Reaction Conditions:

1X SE-Buffer 2K, BSA (100 µg/ml). Incubate at 37° C.

1X SE-Buffer 2K (pH 7.6 @ 25° C):

10 mM Tris-HCl 200 mM KCl 10 mM MgCl₂ 1 mM DTT

Heat Inactivation:

Enzyme is inactivated by incubation at 65°C for 20 minutes.

Unit Definition: One unit is defined as the amount of enzyme required to digest 1 µg of Lambda DNA in 1 hour at 37°C in a total reaction volume of 50 µl. To obtain 100% activity, BSA should be added to the 1 x reaction mix to a final concentration of 100 μg/ml.

Quality Control Assays

Ligation: After 20-fold overdigestion with Ksp22 I, ~90% of the DNA fragments can be ligated and recut.

16-Hour Incubation: A 50 µl reaction containing 1 µg of DNA and 40 Units of enzyme incubated for 16 hours resulted in the same pattern of DNA bands as a reaction incubated for 1 hour.

Do not use BSA for long incubation.

Oligonucleotide Assay: No detectable degradation of a single-stranded and double-stranded oligonucleotide was observed after incubation with 20 units of restriction endonuclease for 3 hours.

Enzyme Properties:

When using a buffer other than the optimal (Supplied) SE-Buffer, it may be necessary to add more enzymes to achieve complete digestion.

Reagents Supplied with Enzyme: 10X SE Buffer 2K, BSA (10mg/ml).

Blocked by overlapping Dam methylation (G^mATC): TGATCA