



Ssp I

Recognition Sequence:

S

500 units 10,000 u/ml

E041

AATĮ ATT TTAT TAA

Lot: Exp:

Store at -20C

 SE-Buffers
 B
 G
 O
 W
 Y
 ROSE

 %Activity
 75-100
 50-75
 25-50
 50-75
 75-100
 100

 55°C
 65°C
 K
 λ
 RR
 BSA

For more details scen the code



CERTIFICATE OF ANALYSIS

<u>Source</u>: An E.coli strain that carries the cloned Sspl gene from Sphaerotilus species.

Supplied in:

10 mM Tris-HCl (pH 7.5), 200 mM NaCl, 0.1 mM EDTA, 1 mM DTT, 200 μg/ml BSA, 50% glycerol.

Reaction Conditions:

1X SE-Buffer K, BSA (100 $\mu g/ml$). Incubate at 37° C.

1X SE-Buffer K (pH 7.6 @ 25° C):

 $\begin{array}{ll} 10~\text{mM Tris-HCl} & 100~\text{mM KCl} \\ 10~\text{mM MgCl}_2 & 1~\text{mM DTT} \end{array}$

Heat Inactivation:

Enzyme is inactivated by incubation at 65°C for 20 minutes.

<u>Unit Definition</u>:One unit is defined as the amount of enzyme required to digest 1 μ g of Lambda DNA in 1 hour at 37° C in a total reaction volume of 50 μ l. To obtain 100% activity, BSA should be added to the 1 x reaction mix to a final concentration of 100 μ g/ml.

Quality Control Assays

<u>Ligation</u>:After 10-fold overdigestion with Ssp I, 90% of the DNA fragments can be ligated and recut.

16-Hour Incubation: A 50 μl reaction containing 1 μg of DNA and 10 Units of enzyme incubated for 16 hours resulted in the same pattern of DNA bands as a reaction incubated for 1 hour.

Do not use BSA for long incubation.

Conditions of high enzyme concentration may result in star activity.

Oligonucleotide Assay:No detectable degradation of a single-stranded and double-stranded oligonucleotide was observed after incubation with 10 units of restriction endonuclease for 3 hours.

Enzyme Properties:

When using a buffer other than the optimal (Supplied) SE-Buffer, it may be necessary to add more enzymes to achieve complete digestion.

Reagents Supplied with Enzyme: 10X SE Buffer K, BSA (10mg/ml).

Blocked by methylation ATATT.