



BspAC I

Recognition Sequence:

E501 200 units 5,000 u/ml CTCCC GGCTG

Lot: Exp:

Store at -20°C

SE-Buffers	В	G	0	w	Υ	ROSE	
%Activity	10-25	25-50	100	75-100	10-25	100	



BSA

For more details scen the code



CERTIFICATE OF ANALYSIS

Source: Bacillus species AC.

Supplied in:

10 mM KH₂PO₄(pH 7.2), 100 mM NaCl, 0.1 mM EDTA, 7 mM 2-mercaptoethanol, 200 µg/ml BSA, 50% glycerol.

Reaction Conditions:

1XSE-Buffer O, BSA (100 µg/ml). Incubate at 37° C.

1X SE-Buffer O (pH 7.6 @ 25° C): 50 mM Tris-HCl 100 mM NaCl 1 mM DTT 10 mM MgCl₂

Heat Inactivation:

Enzyme is inactivated by incubation at 65°C for 20 minutes.

Unit Definition: One unit is defined as the amount of enzyme required to digest 1 µg of Lambda DNA in 1 hour at 37°C in a total reaction volume of 50 µl. To obtain 100% activity, BSA should be added to the 1 x reaction mix to a final concentration of 100 µg/ml.

Quality Control Assays

Ligation: After 5-fold overdigestion with BspAC I, 95% of the DNA fragments can be ligated by using of T4 DNA Ligase and 50% of these can be recut.

16-Hour Incubation: A 50 µl reaction containing 1 µg of DNA and 10 Units of enzyme incubated for 16 hours resulted in the same pattern of DNA bands as a reaction incubated for 1 hour.

Do not use BSA for long incubation.

Oligonucleotide Assay: No detectable degradation of a single-stranded and double-stranded oligonucleotide was observed after incubation with 5 units of restriction endonuclease for 3 hours.

Enzyme Properties:

When using a buffer other than the optimal (Supplied) SE-Buffer, it may be necessary to add more enzymes to achieve complete digestion.

Reagents Supplied with Enzyme: 10X SE Buffer O. BSA (10mg/ml).

Blocked by CG methylation.

BspACI has a non-palindromic recognition site.