

Restriction  
Endonuclease



## Smi I

Recognition  
Sequence:

ATTT↓AAAT  
TAAA↑TTTA

S

E225

1,000 units  
20,000 u/ml

Lot:

Exp:

Store at -20°C

SE-Buffers	B	G	O	W	Y	ROSE
%Activity	25-50	25-50	100	75-100	25-50	10

37°C

65°C

O

T7/Sspl

BSA

For more details  
scan the code



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## CERTIFICATE OF ANALYSIS

Source: *Streptococcus milleri S.*

Supplied in:

10 mM Tris-HCl (pH 7.5), 250 mM NaCl, 0.1 mM EDTA,  
7 mM 2-mercaptoethanol, 100 µg/ml BSA, 50%  
glycerol.

Reaction Conditions:

1X SE-Buffer O, BSA (100 µg/ml). Incubate at 37° C.

1X SE-Buffer O (pH 7.6 @ 25° C):

50 mM Tris-HCl    100 mM NaCl  
10 mM MgCl<sub>2</sub>    1 mM DTT

Heat Inactivation:

Enzyme is inactivated by incubation at 65°C for 20  
minutes.

Unit Definition: One unit is defined as the amount of  
enzyme required to digest 1 µg of T7 DNA/Sspl in 1  
hour at 37° C in a total reaction volume of 50 µl. To  
obtain 100% activity, BSA should be added to the 1  
x reaction mix to a final concentration of 100 µg/ml.

Quality Control Assays

Ligation: After 20-fold overdigestion with Smi I, > 95%  
of the DNA fragments can be ligated by using of high  
concentration T4 DNA Ligase with 10% PEG and recut.

16-Hour Incubation: A 50 µl reaction containing 1 µg of  
T7 DNA and 40 Units of enzyme incubated for 16 hours  
resulted in the same pattern of DNA bands as a reaction  
incubated for 1 hour.

Do not use BSA for long incubation.

Oligonucleotide Assay: No detectable degradation of a  
single-stranded and double-stranded oligonucleotide  
was observed after incubation with 20 units of restriction  
endonuclease for 3 hours.

Enzyme Properties:

When using a buffer other than the optimal (Supplied)  
SE-Buffer, it may be necessary to add more enzymes  
to achieve complete digestion.

Reagents Supplied with Enzyme:

10X SE Buffer O, BSA (10mg/ml).