

Restriction  
Endonuclease



# AccB1 I

Recognition  
Sequence:

G↓GYRCC  
CCRYG↑G

S

**E163**

500 units  
5,000 u/ml

Lot:

Exp:

Store at -20°C

SE-Buffers	B	G	O	W	Y	ROSE
%Activity	50-75	10-25	10-25	75-100	50-75	30

37°C

65°C

K

λ

BSA

For more details  
scan the code



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## CERTIFICATE OF ANALYSIS

Source: *Acinetobacter calcoaceticus B1*.

Supplied in:

10 mM Tris-HCl (pH 7.5), 50 mM KCl, 0.1 mM EDTA,  
7 mM 2-mercaptoethanol, 200 µg/ml BSA, 50%  
glycerol.

Reaction Conditions:

1x SE-Buffer K, BSA (100 µg/ml). Incubate at 37° C.

1X SE-Buffer K (pH 7.6 @ 25° C):

10 mM Tris-HCl    100 mM Kcl  
10 mM MgCl<sub>2</sub>    1 mM DTT

Heat Inactivation:

Enzyme is inactivated by incubation at 65 °C for 20  
minutes.

Unit Definition: One unit is defined as the amount of  
enzyme required to digest 1 µg of λ DNA in 1 hour at  
37° C in a total reaction volume of 50 µl.

To obtain 100% activity, BSA should be added to the  
1x reaction mix to a final concentration of 100 µg/ml.

Quality Control Assays

Ligation: After 5-fold overdigestion with AccB1 I, 95%  
of the DNA fragments can be ligated with T4 DNA Ligase  
and recut.

16-Hour Incubation: A 50 µl reaction containing 1 µg of  
DNA and 5 Units of enzyme incubated for 16 hours  
resulted in the same pattern of DNA bands as a reaction  
incubated for 1 hour.

No using BSA for long incubation.

High enzyme concentration results in star activity.

Oligonucleotide Assay: No detectable degradation of a  
single-stranded and double-stranded oligonucleotide  
was observed after incubation with 5 units of restriction  
endonuclease for 3 hours.

Enzyme Properties:

When using a buffer other than the optimal (Supplied)  
SE-Buffer, it may be necessary to add more enzymes  
to achieve complete digestion.

Reagents Supplied with Enzyme:

10X SE Buffer K, BSA (10 mg/ml).